Statistical analysis of absorption spectra of phytoplankton and of pigment concentrations observed during three POMME cruises using a neural network clustering method

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We present a neural network methodology for clustering large data sets into pertinent groups. We applied this methodology to analyze the phytoplankton absorption spectra data gathered by the Laboratoire d’Océanographie de Villefranche. We first partitioned the data into 100 classes by means of a self-organizing map (SOM) and then we clustered these classes into 6 significant groups. We focused our analysis on three POMME campaigns. We were able to interpret the absorption spectra of the samples taken in the first oceanic optical layer during these campaigns, in terms of seasonal variability. We showed that spectra from the PROSOPE Mediterranean campaign, which was conducted in a different region, were strongly similar to those of the POMME-3 campaign. This analysis led us to propose regional empirical relationships, linking phytoplankton absorption spectra to pigment concentrations, that perform better than the previously derived overall relation. ©2007 Optical Society of America

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1. Introduction
The principal objective of analyzing large databases is to extract pertinent information, such as seasonal and regional characteristics [1,2,3] and to present this information in a suitable form to facilitate its interpretation. Recently Chazottes et al. [4] proposed an advanced neural network method for analyzing the Laboratoire d’Océanographie de Villefranche (LOV) data set which is a large set of phytoplanktonic absorption spectra. In the present paper we present a methodology derived from that of Chazottes et al. [4] and able to capture regional and seasonal information embedded in the same data set.

The LOV has gathered a large set of ocean water samples for which the absorption spectra of phytoplankton and the corresponding pigment concentrations have been measured. These samples were taken during several cruises covering different parts of the world ocean at different seasons and therefore present a wide variety of situations. Chazottes et al. [4] processed the phytoplankton absorption spectra with a sophisticated neural network method suitable for classifying complex phenomena, the so-called self-organizing map (SOM) proposed by Kohonen [5]. The aim was to compress the information embedded in the data set into a reduced number of classes, the so-called reference vectors, rv, which characterize the data set; this facilitates the subsequent analysis. Chazottes et al. [4] were thus able to retrieve well-known relationships among pigment concentrations and to display them on maps to facilitate their interpretation. They were also able to propose new empirical relationships linking absorption spectra and pigment concentrations. In Chazottes et al. [4], the
number of classes is quite large (10 × 10), making some analyses, such as that of spatial or temporal variability, quite difficult. The objective of the present paper is to extend the work of Chazottes et al. [4] by clustering the SOM classes into a small number of groups in order to facilitate the interpretation in terms of bio-optical considerations. We were able to reveal regional and seasonal structures for which specific empirical relationships between absorption spectra and pigment concentrations can be proposed for each group. Focus is placed on the analysis of the biogeochemical characteristics of several oceanic regions represented in the LOV database.

2. Data and Methods
The database we processed and the SOM algorithm have been fully described by Chazottes et al. [4]; we recall here its major characteristics.

A. Database and Data Sets
Water samples were collected during ten cruises, in various seasons and various areas of the world ocean, between 1990 and 2001 (Table 1). In this study, we consider phytoplankton absorption spectra as pigment concentrations corresponding exclusively to oceanic case-1 waters.

Methods employed for particulate and algal absorption measurements are described in detail by Bricaud et al. [6,7]. The absorption spectrum was measured every 2 nm from 400 to 700 nm. We then applied a triangular moving window of size 3 to filter the noisy part of each spectrum. The filtered spectra were then sampled every 10 nm. Each spectrum is therefore represented by a 31-dimension vector. The phytoplankton spectral absorption coefficients are represented by the symbol $a_{ph}(\lambda)$ where $\lambda$ stands for the wavelength in nanometers (nm).

Pigment concentrations were measured by high-pressure liquid chromatography (HPLC), using the procedure described by Vidussi et al. [8]. All the pigments were grouped into five main categories, according to their spectral similarities [6,7].

Owing to the large variation (covering several decades) in the absorption spectral values, $a_{ph}(\lambda)$, we subsequently used $\log_{10}(a_{ph}(\lambda))$ values rather than $a_{ph}(\lambda)$ values, and for analogous reasons we also used the log-transformed values of the pigment concentrations.

Each absorption spectrum is thus represented by a 31-component vector, whose first 30 components are the spectrum derivatives computed as the difference between the $\log_{10}(a_{ph}(\lambda))$ values for two consecutive wavelengths (i.e., $\log(a_{ph}(\lambda_{400+10})) - \log(a_{ph}(\lambda_{400+10}+1))$, where $i = 1 \ldots 30$), and the last component is the maximum value of the absorption [4].

The whole data set, $D$, which is described in Table 1, comprises 3734 samples. In the present study the learning data set, $L$, which was used to estimate the parameters of the self-organizing map, comprised 2163 samples. These data are from the samples of various cruises, among which are those of the POMME 1, 2, and 3 cruises. To avoid a possible bias due to the large number of the POMME data, only 525 POMME samples were retained in the learning set, $L$. The remaining 1571 POMME samples constituted a validation set, denoted hereinafter $V$.

In the present study, we focused on the properties of water samples in the surface layer, the most accessible to ocean color sensors and the most active one in terms of seasonal biological activity. The “surface” layer has been defined here as having a thickness equal to the penetration depth (i.e., the depth above which 90% of the diffusely reflected irradiance originates), computed with respect to the photosynthetically available radiation (PAR). This penetration depth is approximately equal to $\rho_{e}/4.6$, where $\rho_{e}$ represents the euphotic depth. This euphotic depth was either directly measured during the different cruises or computed from the chlorophyll profile according to Morel and Maritorena [9].

### Table 1. Information Concerning the Cruises on which the Different Water Samples Were Collected

<table>
<thead>
<tr>
<th>Cruises</th>
<th>Location</th>
<th>Usual Trophic State</th>
<th>Date</th>
<th>Number of Samples</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 TOMOFRONT</td>
<td>Northwestern Mediterranean</td>
<td>Mesotrophic</td>
<td>April 1990</td>
<td>28</td>
</tr>
<tr>
<td>2 EUMELI3</td>
<td>Tropical North Atlantic</td>
<td>Oligotrophic, mesotrophic</td>
<td>Oct. 1991</td>
<td>49</td>
</tr>
<tr>
<td>3 FLUPAC</td>
<td>Equatorial and subequatorial Pacific</td>
<td>Oligotrophic</td>
<td>Sept.–Oct. 1994</td>
<td>80</td>
</tr>
<tr>
<td>4 OLIPAC</td>
<td>Equatorial and subequatorial Pacific</td>
<td>Oligotrophic</td>
<td>Nov. 1994</td>
<td>183</td>
</tr>
<tr>
<td>5 MINOS</td>
<td>Eastern and western Mediterranean</td>
<td>Oligotrophic</td>
<td>May 1996</td>
<td>115</td>
</tr>
<tr>
<td>8 PROSOPE (upw)</td>
<td>Morocco upwelling area</td>
<td>Eutrophic</td>
<td>September 1999</td>
<td>52</td>
</tr>
<tr>
<td>9 POMME 1</td>
<td>North Atlantic</td>
<td>Mesotrophic</td>
<td>Feb.–March 2001</td>
<td>187 + (561)</td>
</tr>
<tr>
<td>10 POMME 2</td>
<td>North Atlantic</td>
<td>Mesotrophic</td>
<td>March–May 2001</td>
<td>193 + (577)</td>
</tr>
<tr>
<td>12 BENCAL</td>
<td>Benguela upwelling area</td>
<td>Mesotrophic, eutrophic</td>
<td>Oct. 2002</td>
<td>100</td>
</tr>
</tbody>
</table>

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We note that the SOM associated with the neurons of the SOM is given in Fig. 1. The resulting clustering of the spectra as shown). Figure 2 shows the upper part of the dendrogram in which groups 7 and 8 have been left apart, owing to the low number of particular spectra that fell into them. Two groups connected by the same dendrite present more similarities than those connected by hierarchical dendrites.

In the following, we study the variability of the phytoplankton absorption spectra from samples corresponding only to the first optical depth (surface waters), which are those observed by satellite sensors. For each group, we computed the mean and standard deviation corresponding to the spectra retained by its neurons, the mean values of the spectrum derivatives (as defined previously), the mean pigment concentrations and their normalized ratios (i.e. pigment concentration:Tchl-a ratios). In Chazottes et al. [4], the neurons, which were clustered according to their Tchl-a concentration, were also found to be associated with pigment ratios relative to Tchl-a (Tchl-b/Tchl-a, etc.). Although Fig. 3 shows that the groups are ranked in decreasing order with respect to their Tchl-a concentration, the various group ranges partially overlap each other. Figure 4 displays the mean pigment concentrations relative to Tchl-a of the L data set for each group. We note the different patterns of groups 4, 5, and 6 with respect to that of groups 1, 2, 3 which present a lower TPPC/Tchl-a ratio, showing that SOM + HAC is able to cluster the water samples not only from the amplitude of their absorption spectra, but also from their

Table 2. Sample Repartition of the Surface Samples in the Different Data Sets and Groups

<table>
<thead>
<tr>
<th>Learning Set: Lsurf</th>
<th>Validation Set: Vsurf</th>
</tr>
</thead>
<tbody>
<tr>
<td>Learning</td>
<td>PROSOP-E-Med</td>
</tr>
<tr>
<td>Group 1</td>
<td>50</td>
</tr>
<tr>
<td>Group 2</td>
<td>84</td>
</tr>
<tr>
<td>Group 3</td>
<td>24</td>
</tr>
<tr>
<td>Group 4</td>
<td>62</td>
</tr>
<tr>
<td>Group 5</td>
<td>141</td>
</tr>
<tr>
<td>Group 6</td>
<td>16</td>
</tr>
<tr>
<td>All groups</td>
<td>377</td>
</tr>
</tbody>
</table>

Fig. 1. Clustering of the neurons on the SOM. Following the HAC, based on the maximum amplitude and the slopes of the spectra, eight groups were retained. The group number resulting from the HAC is displayed for each neuron. The SOM + HAC clustering is coherent, since the groups represent clusters of contiguous neurons.

Fig. 2. Top of the dendrogram resulting from the HAC is presented for the six groups used in the present study. The number of data of L corresponding to each group is also known. The higher the node linking two groups the further apart they are in the data space.
pigment composition. In particular, Fig. 5 shows the partition of the $L_{surf}$ samples for the different cruises in the six groups that cluster samples having similar spectral characteristics. We note that groups 2, 3, 4, 5, 6 gather samples coming from at least three cruises corresponding to different oceanic regions and different seasons.

D. Interest of the Method

Several questions may be raised on the above methodology. The first one concerns its usefulness. Could the following analysis have been conducted with a careful inspection of the database? The answer is yes, but with a lot of effort and laborious comparisons and tests. A major advantage of the SOM + HAC method is its highly efficient discrimination in producing a coherent first-order classification of the absorption spectra (amplitude of the spectra associated with the Tchl-a) as well as a second-order classification (pigment concentrations normalized by Tchl-a concentration i.e. pigmentary composition). A second question is the visualization associated with the SOM and the group clustering, allowing us to easily and quickly capture specific information embedded in the database.

Two theoretical questions remain concerning the HAC. First, could we have carried out the HAC directly on the data instead of on the $r_v$ of the SOM? It has been shown [12] that it is more efficient to do the HAC on the $r_v$, which represent the most significant synthetic observations associated with the database than to apply it directly to the data which may be noisy. We could also have directly used a $(3 \times 2)$ SOM to partition the full data set into a small number of clusters. We tested such a SOM algorithm. The partition we obtained is less pertinent in terms of physical interpretation than the SOM + HAC method when looking at the different elements of the database clustered in the different classes. The reason is that the $(3 \times 2)$ SOM does not have a sufficient degree of liberty with respect to the $(10 \times 10)$ SOM + HAC algorithm to adequately extract the statistical information from the original database that can be noisy and to make a pertinent partition.

In the following we used SOM + HAC to analyze the POMME samples of the surface ($V_{surf}$) and some bio-optical relationships relating pigment concentrations to absorption spectra.

3. Analysis of Absorption Spectra from the POMME Cruises

We now try to characterize the biogeochemical variability of the POMME region by analyzing the phytoplanktonic absorption spectra with the SOM + HAC method. The POMME (Programme Océan Multidisciplinaire Meso Echelle) experiment took place in the North Atlantic Ocean (21.33°W, 38.00°N) [13]; the region was extensively sampled between October 2000 and September 2001.

We processed the phytoplankton absorption spectra from the 217 $V_{surf}$ surface samples from the
POMME 1, 2, and 3 cruises, using the SOM + HAC method described in Section 2. Table 2 shows the distribution of the data among the various groups for the learning surface data set $L_{surf}$ and the three POMME campaigns ($V_{surf}$). In Fig. 6, for each of the six groups, we show the mean and the standard deviation of the spectra (estimated using $L_{surf}$) as well as the mean spectra for each of the three POMME cruises (val-P1, val-P2 and val-P3) and for the PROSOPE-Med cruise (learn-Pmed) have been superposed.

In the following we used the SOM + HAC visualization in order to analyze the result of the classification for each campaign.

The amplitude of the mean absorption spectrum of POMME-1 is lower (Fig. 6) than that of group 2 surface water samples. This is in agreement with the low mean pigment concentrations of POMME-1 with respect to that of surface waters of group 2, as presented in Fig. 8. In Fig. 9, we plotted the four pigment concentrations relative to Tchl-a for the six groups. The POMME-1 Tchl-b/Tchl-a ratio is particularly high compared to that of the other groups, in agreement with Bricaud et al. [14]. Figure 10 shows that the amplitude of the mean group 2 absorption spectrum for the full learning data set, $L$, is smaller than that of the $L_{surf}$ data set and close to that for POMME-1.

We should recall that the POMME-1 cruise was conducted from January to March, just before the phytoplankton spring bloom [13,15]. During that period the depth of the mixed layer exceeded 120 m [13,16]. This led us to tentatively interpret the behavior described above according to the following scenario: since the mixed layer was deep, the POMME-1 phytoplankton species are probably deep species, as shown by the high Tchl-b/Tchl-a ratio. (It should be noted that the Tchl-b index also contains divinyl-
chlorophyll-b which is the indicator of prochlorophytes, a not necessarily deep species.) The low amplitude of the absorption spectrum and the pigment concentration is due to the fact that POMME-1 was conducted before the spring bloom, when the phytoplankton concentration was low.

The POMME-2 cruise was conducted from March to May, just after the spring bloom. The mixed layer during that period was much shallower than during POMME-1. The fact that the POMME-1 and POMME-2 spectra are both clustered in group 2 shows that the POMME-2 spectrum is similar to that of POMME-1 (Fig. 6). This is confirmed by the spectrum derivatives that are both closely related to the group 2 reference vector (Fig. 7). The similarity between the POMME-1 and the POMME-2 spectra and spectrum derivatives suggests that the POMME-2 phytoplankton results from the blooming of the POMME-1 phytoplankton population. The major difference between the two spectra and the two pigment means is the amplitude difference as seen in Figs. 6 and 8. This is in agreement with a higher phytoplankton concentration observed during the POMME-2 cruise than during POMME-1.

The POMME-3 cruise took place in September and October. The POMME-3 mean spectrum, which mainly belongs to group 5 (Table 2), is very different from those of POMME-1 and POMME-2 (Fig. 6 for the amplitude; Fig. 7 for the derivatives). Furthermore, the POMME-3 normalized pigment concentrations (Fig. 9), being also very different from those of POMME-1 and POMME-2, we can argue that the phytoplankton species were different during POMME-3. In fact, the POMME-3 cruise was held a sufficient time after the spring bloom, which apparently occurred between the POMME-1 and the POMME-2 cruises, to be uncorrelated with them [13,15].

Since SOM + HAC allows us a pertinent biogeo-physical interpretation of results from new cruises whose samples were not included in the learning set, we propose to use the clustering into 6 groups to give a regionalized account of various relations linking phytoplankton absorption spectra and pigment concentrations.

4. Determination of Regional Bio-optical Relations

POMME-3 and PROSOPE Mediterranean surface data were captured by the same neurons of the SOM that belong to the group 5 (Table 2). This is due to the fact that the mean POMME-3 and the PROSOPE phytoplankton absorption spectra (Fig. 6) are close to that of the mean group 5, suggesting that the absorption properties of the phytoplankton in the POMME-3 and PROSOPE Mediterranean surface waters were very similar. The pigment values and their normalized ratio (Figs. 8 and 9) were also closely related.

Based on this affirmation, we considered the 6 groups as a regionalization in the data space which is expected to provide better-fitted relationships linking phytoplankton absorption spectra and pigment concentrations.

<table>
<thead>
<tr>
<th>Group</th>
<th>Intercept</th>
<th>Slope</th>
<th>R²</th>
<th>s²</th>
<th>rmse</th>
<th>s²</th>
<th>rmse</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>-1.121</td>
<td>0.604</td>
<td>0.731</td>
<td>0.108</td>
<td>0.106</td>
<td>0.115</td>
<td>0.113</td>
</tr>
<tr>
<td>2</td>
<td>-1.204</td>
<td>0.465</td>
<td>0.438</td>
<td>0.112</td>
<td>0.112</td>
<td>0.131</td>
<td>0.130</td>
</tr>
<tr>
<td>3</td>
<td>-1.470</td>
<td>0.327</td>
<td>0.721</td>
<td>0.051</td>
<td>0.049</td>
<td>0.156</td>
<td>0.151</td>
</tr>
<tr>
<td>4</td>
<td>-1.303</td>
<td>0.637</td>
<td>0.753</td>
<td>0.084</td>
<td>0.083</td>
<td>0.100</td>
<td>0.099</td>
</tr>
<tr>
<td>5</td>
<td>-1.315</td>
<td>0.647</td>
<td>0.770</td>
<td>0.072</td>
<td>0.072</td>
<td>0.083</td>
<td>0.083</td>
</tr>
<tr>
<td>6</td>
<td>-2.169</td>
<td>0.176</td>
<td>0.046</td>
<td>0.091</td>
<td>0.085</td>
<td>0.243</td>
<td>0.227</td>
</tr>
</tbody>
</table>

Fig. 9. Group clustering of the mean normalized pigment concentration in samples from the first optical layer. The mean normalized pigment concentrations of the samples from the three POMME cruises (val-P1, val-P2 and val-P3) and the Prosope-Med cruise (learn-Pmed) are displayed. We have also displayed the mean normalized pigment concentrations and their standard deviation computed from the learning data set for each group (open circles).

Fig. 10. Mean absorption spectrum of the group 2 surface waters (o), of the group 2 water column including deep and surface waters, (V), of the POMME-1 (x) and POMME-2 (+) cruises. The POMME-1 spectrum is close to the mean spectrum of the group 2 which includes deep samples.
centrations at regional scale. Indeed, initially, the concept behind regionalization was to adapt the parameters of some algorithm to a specific region of the world ocean. Recently, Alvain et al. [17] proposed a “regionalization” of the OC4V4 SeaWiFS algorithm based on classes of normalized radiance. Similarly, our regionalization is based on resemblances of phytoplankton absorption spectra linked to pigment concentrations. In the following subsection we revisit some absorption:concentration relationships using it.

A. Relation between \( a_{ph}(440) \) and Tchl-a

Bricaud et al. [14] proposed a widely used relationship allowing computation of the phytoplankton absorption coefficient at 440 nm from the Tchl-a concentration (for the first optical depth waters), which is of the form

\[
a_{ph}(440) = 0.0654Tchl-a^{0.728}
\]

(with \( R^2 = 0.934, N = 596 \)).

We estimated a similar power-law relationship for each SOM + HAC group. The regression lines for the 6 groups are presented in Fig. 11, concomitant with the Bricaud et al. [14] regression line. The group

<table>
<thead>
<tr>
<th>HAC</th>
<th>Bricaud et al. [14]</th>
</tr>
</thead>
<tbody>
<tr>
<td>( R^2 )</td>
<td>( s^2 )</td>
</tr>
<tr>
<td>0.961</td>
<td>0.092</td>
</tr>
</tbody>
</table>

Table 4. Global (on the Whole Data Set) Performance Evaluation for the SOM + HAC Regression Lines and for the Bricaud et al. [14] Regression Line

Fig. 11. (a) Scatterplot of the Bricaud et al. [14] regression and (b)–(g) Bricaud et al. [14] and HAC-group relationships linking absorption to Tchl-a for the six groups.
regression lines better fit the data than the Bricaud et al. [14] regression does. This is confirmed by the fact that the rms computed for each group regression line is much smaller than that computed using the Bricaud et al. [14] regression line (Tables 3 and 4). These six regressions constitute a piecewise fitting of the relationship. The $R^2$ estimator of this piecewise fitting was 0.961, which is larger than the corresponding Bricaud et al. [14] value of 0.934, showing that the Bricaud et al. [14] relationship is valid at the first-order level but is unable to fit second-order non-linearity. Furthermore, Fig. 11 clearly shows that the group 2 relationship, which has a wavelike shape, is not a power law and is governed by a more complex relationship. The partition of the data set into groups allowed us to refine the exploration and the understanding of the behavior of that data set.

B. Relation between the Derivative of $a_{\text{pr}}(640)$ and the Tchl-b/Tchl-a Ratio

Chazottes et al. [4] proposed a new empirical relationship between the Tchl-b/Tchl-a ratio and the derivative of the absorption spectrum at 640 nm (whenever Tchl-b is not zero). This relationship has been estimated on the global learning data set $L$. It is of the form

\[
\text{Tchl-b/Tchl-a} = 0.090 \left(\frac{a_{650}}{a_{640}}\right)^{0.836}
\]

(with $R^2 = 0.531$ and $s = 0.246$). \(2\)

We have computed a similar relationship using the samples from the first optical layer. It is of the form

\[
\text{Tchl-b/Tchl-a} = 0.0684 \left(\frac{a_{650}}{a_{640}}\right)^{0.624}
\]

(with $R^2 = 0.431$ and $s = 0.240$). \(2'\)

We estimated similar relationships calibrated for the surface samples of each group. The objective was to obtain specific regional regressions that should be more accurate in each group than the overall regression. The specific regressions are much better than the overall regression for groups 1 and 2 ($R^2 = 0.656$ and $s = 0.174$ and $R^2 = 0.747$ and $s = 0.142$, respectively). The goodness of fit drops dramatically for the other groups ($R^2 = 0.36$ for group 3, $R^2 = 0.38$ for group 4, $R^2 = 0.26$ for group 5 and $R^2 = 0.002$ for group 6). This means that the relationship \(2'\) is mainly driven by data belonging to groups 1 and 2. The goodness of fit decreases with the group number. This can be due to the fact that the pigment concentrations strongly decrease with the group number, the relation is valid only above a given concentration threshold.

As mentioned above, POMME-1 and POMME-2 absorption spectra belong to group 2, while POMME-3 spectra belong to group 5. Figure 12 shows the new relationships calibrated on the group 2 $L_{\text{surf}}$ data set and on the group 2 surface samples, respectively. We note that the group 2 (POMME-1 and POMME-2) data better fit the new regression calibrated on group 2 surface data than the regression line calibrated on $L_{\text{surf}}$.

C. Relationship between the Derivative of $a_{\text{pr}}(500)$ and the Fucoxanthin/Tchl-a Ratio

We did a similar study for the fucoxanthin/Tchl-a ratio and the derivative of the absorption spectrum at 510 nm. As in the previous section, we reestimated the regression proposed by Chazottes et al. [4] on the samples in the first optical layer ($L_{\text{surf}}$), which is of the form

\[
\text{Fucoxanthin/Tchl-a} = 1.311 \left(\frac{a_{510}}{a_{500}}\right)^{0.74}
\]

(with $R^2 = 0.46$, $s = 0.264$). \(3\)

We estimated a similar power-law relationship for each SOM + HAC group, using its specific surface data. Only the group 2 regression presents a better goodness of fit ($R^2 = 0.685$ and $s = 0.193$) than that estimated from the learning surface data set, $L_{\text{surf}}$. Figure 13, which is similar to Fig. 12, shows the new relationships, together with the surface data from the POMME-1 and POMME-2 cruises. The POMME-1 and POMME-2 surface data better fit the new regression calibrated on group 2 surface data than the re-
We revisited the empirical relationship proposed by Briceaud et al. [14] linking the absorption at 440 nm to Tchl-a by computing a specific regression for each group. These different regressions constituted a piecewise fitting of the relationship, which better fits the data than does the Briceaud et al. [14] regression, showing that there exists some second-order nonlinearity which the Briceaud et al. [14] regression does not account for. This nonlinearity is evident for the group 2 data presented in Fig. 11.

We also revisited the Chazottes et al. [4] relationships linking the Tchl-b/Tchl-a ratio to the derivative of the absorption spectrum at 640 nm and the fucoxanthin/Tchl-a ratio to the derivative of the absorption spectrum at 510 nm. The aim was to propose an improved specific relationship for each group. We found that these two relationships were significantly better for group 2 data only and that there was a somewhat improved relationship for the Tchl-b/Tchl-a ratio for group-1 data. Interpretation may be sought in the decrease in the pigment concentrations as the group number increases and consequently an implicit threshold effect on the pigment concentration, the relationships being valid above a certain concentration only. Another reason might be the phytoplankton diversity associated with the different groups as shown by the analysis presented in Appendix A.

Appendix A

We divided the samples into three water types according to Tchl-a concentrations, based on their Tchl-a concentrations. For the sake of simplification, we will hereafter call “oligotrophic” those waters with [Tchl-a] < 0.2 mg.m$^{-3}$, “mesotrophic” those with [Tchl-a] between 0.2 mg.m$^{-3}$ and 1 mg.m$^{-3}$, and “eutrophic” those with [Tchl-a] > 1 mg.m$^{-3}$. Applying the SOM-HAC algorithm gave the number of samples, for each water type, captured by each group: group 1 contained eutrophic waters only; group 2, and 3 were predominantly mesotrophic waters. These three groups are contiguous on the SOM (Fig. 14). Group 4 comprised a mixture of mesotrophic and oligotrophic waters, and groups 5 and 6 contained oligotrophic water only.

In Fig. 4, the normalized total photosynthetic carotenoids (TPSCs) are much larger than the nor-

![Fig. 14. Histogram of the distribution of the surface-water samples from the water types in each of the six groups. The histogram is computed as the ratio of the number of samples of a given water type in a given group to the total number of samples in that group. The number of water samples is given for each group. 1, 2, and 3 stand for the different water types, respectively, oligotrophic, mesotrophic, and eutrophic.](image-url)
normalized total photoprotectant carotenoids (TPPCs) for the first three groups (1, 2, 3), which is a characteristic of eutrophic and mesotrophic surface waters; the contrary holds for the other three groups (4, 5, 6) and is a characteristic of deep oligotrophic waters. Besides, the group-1 data came mainly from the Bencal campaign, which was conducted in a highly productive area with typical phytoplankton species. Group 2 data, which came from several different campaigns (Fig. 5) are mainly associated with mesotrophic waters (Fig. 14), whereas groups 4, 5, and 6 waters are oligotrophic. The mean absorption spectrum of group 2 being very different in amplitude and shape from those of groups 4, 5, and 6, we may argue that the phytoplankton composition is quite different from that of groups 4, 5, and 6.

Therefore the SOM + HAC groups correspond to a water type and to a mean pigmentary composition.

Appendix B: Linear Regression

To estimate the quality of a linear or log-linear relationship, two parameters, $R^2$ and $s$, are usually calculated. Let us consider a data set of $n$ samples $(x_i, y_i)^{\text{obs}}$. The determination of the parameter $R^2$, which is a measure of “goodness of fit,” represents the part of the variation in $y$ explained by $x$. It is given by

$$R^2 = \frac{\sum (y_i^{\text{estimated}} - \bar{y})^2}{\sum (y_i^{\text{observed}} - \bar{y})^2}, \quad (B1)$$

where $\bar{y}$ is the mean of the $y_i^{\text{observed}}$. The parameter $s$, which is referred to as the root mean square (rms), is an estimate of the error on $y$. It is given by

$$s = \sqrt{\frac{\sum (y_i^{\text{observed}} - y_i^{\text{estimated}})^2}{n - 2}}. \quad (B2)$$

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